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# You're in the right place.

Read more about Teiko's spectral flow service @ teiko.bio/spectral-flow-cytometry

### Fresh versus fixed comparison 25-marker spectral flow cytometry

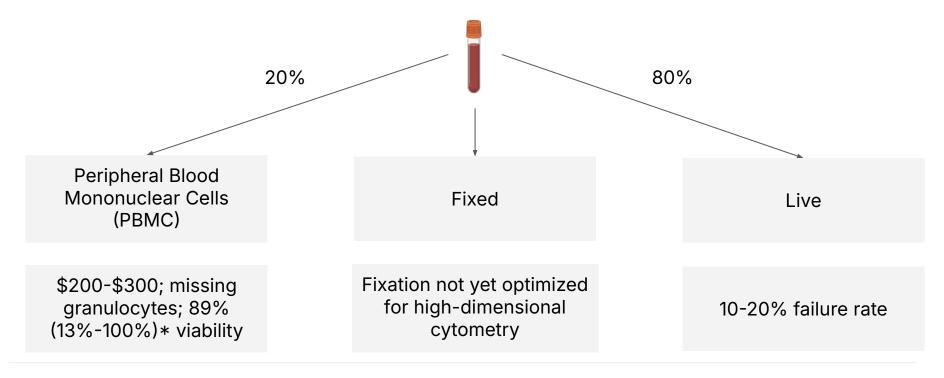
High-parameter cytometry for clinical trials



Ramji Srinivasan Teiko CEO

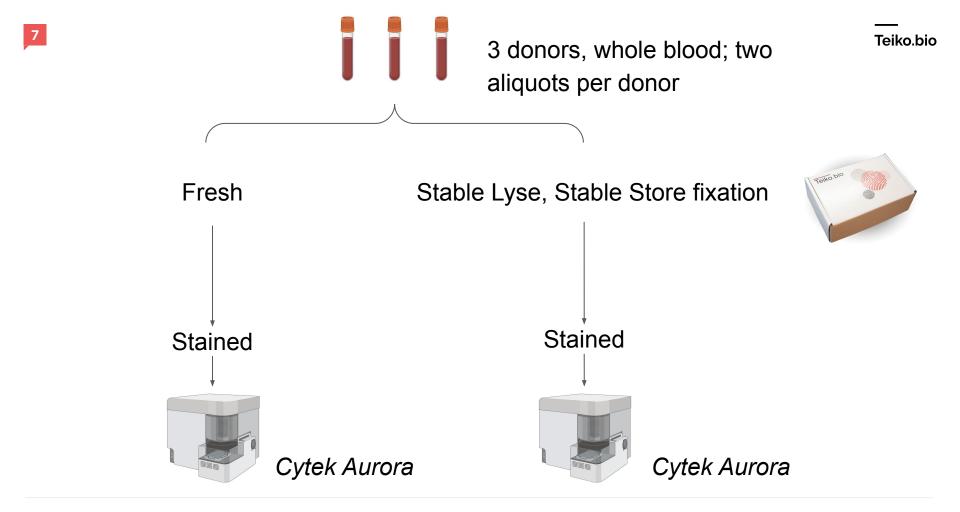
# Problem

#### The drug developer's dilemma

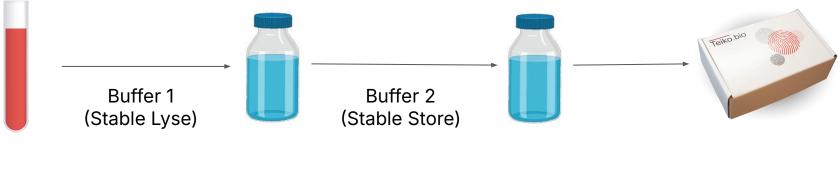


### Does a fixed whole blood sample look similar to a fresh one run on spectral flow?

### Method



#### **TokuKit collection process**

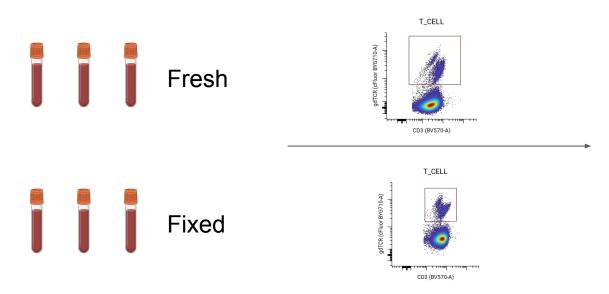


"Pre-fixation"

Target	Fluorophore	Fresh	Fixed
CD66b	Spark UV® 387	X*	$\checkmark$
CD25	BUV661		
CD127	BUV737		$\checkmark$
CD3	BV570		
CD28	BV650		$\checkmark$
CD38	BV711		
CD56	BV750		$\checkmark$
lgM	BV510		
PD-1	BV785		$\checkmark$
CD45RA	cFluor® V450		
CD20	cFluor® V547		$\checkmark$
CD141	cFluor® B515		
CD8	cFluor® B532		$\checkmark$
CD14	cFluor® B548		
HLA-DR	cFluor® B690		$\checkmark$
CD16	cFluor® BYG610		
lgD	cFluor® BYG667		$\checkmark$
CD4	cFluor® YG584		
CD11c	cFluor® BYG781		$\checkmark$
CD1c	cFluor® R668		
CD19	cFluor R685		$\checkmark$
CD123	cFluor R720		
CD45	cFluor R780		$\checkmark$
CD27	cFluor R840		
gdTCR	cFluor BYG710		$\checkmark$

\* Neutrophil exclusion on fresh samples is done via scatter plot. During the fixation protocol, side scatter information is largely lost. Therefore we use the common neutrophil marker CD66b on fixed samples to exclude neutrophils.

#### What we compared



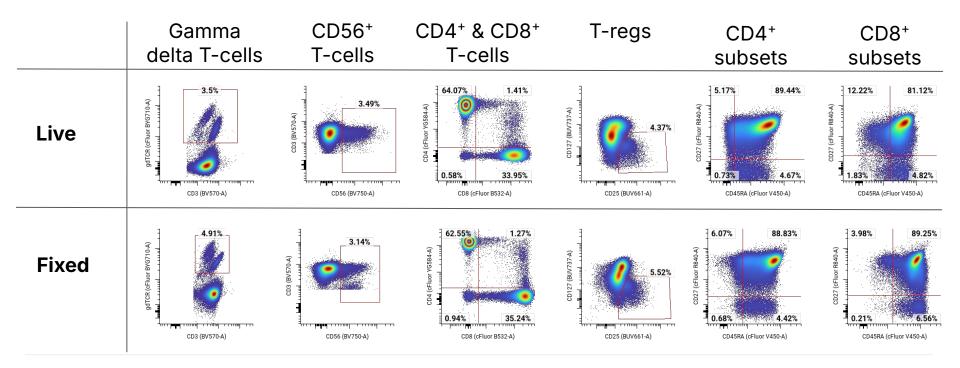
Compared 36 phenotypic populations, and 62 state marker subtypes (i.e. PD-1 in CD4)

Measures: correlation of population frequencies between fresh and fixed

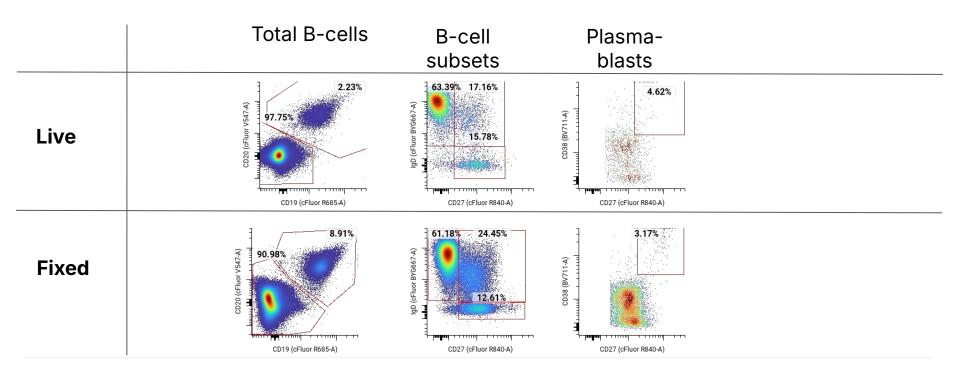
# Results

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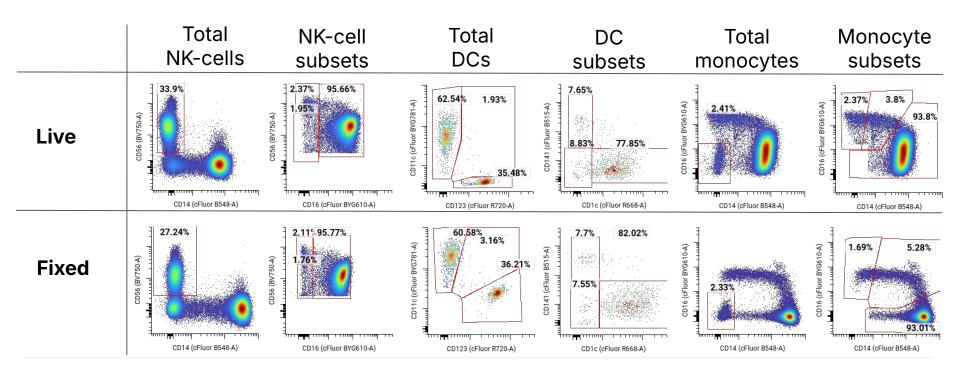
### How different do plots look like from samples stained live versus fixed? Let's start with T cells.



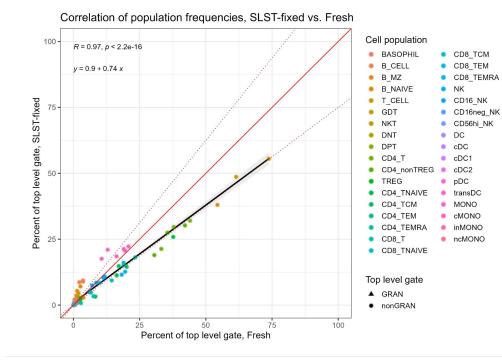
#### How about B cells?



#### NK? DCs? And monocytes?

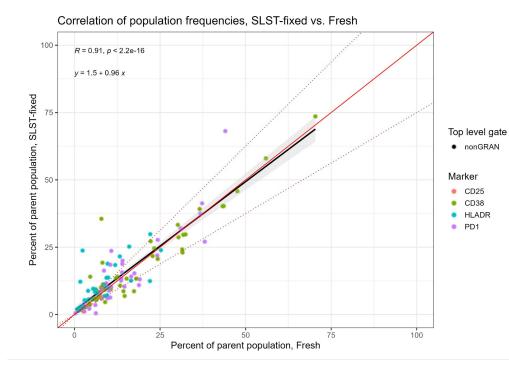


#### Overall, how do the two approaches correlate?



Overall correlation of population frequencies between live and fixed sample processing: **0.97** 

#### What about state markers?

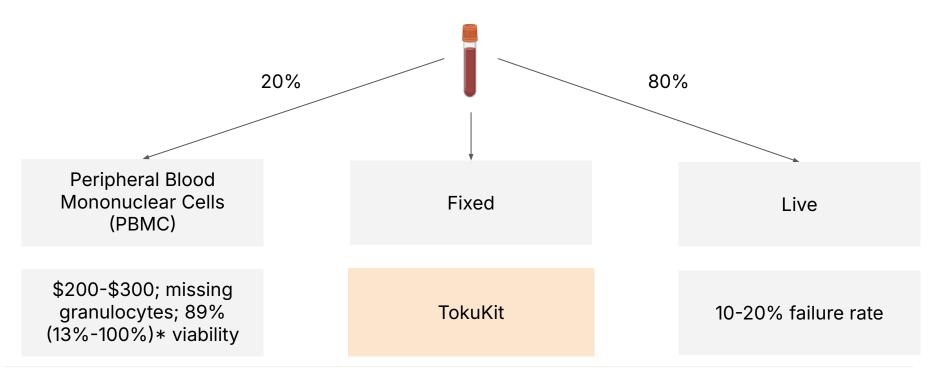


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Overall correlation of state marker-positive subset frequencies between live and fixed sample processing: **0.91** 

## In sum

#### The drug developer's dilemma resolved



### Does a fixed whole blood sample look similar to a fresh one run on spectral flow?

# What this enables

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#### 1. 10-20X reduction of failure rate through centralization of flow analysis No need for a flow lab in every country, city, clinical site. Reduce variability by having all samples processed by the same facility.

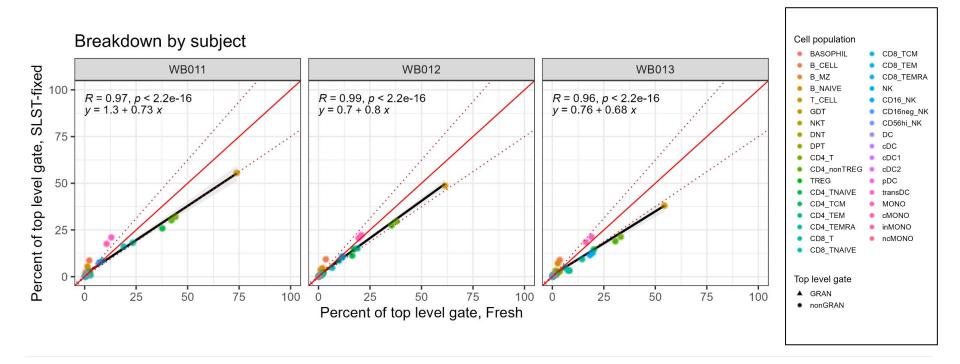
#### 2. Enables batching of samples

Conveniently store fixed samples until a whole set (for example all timepoints of an individual patient) is complete.

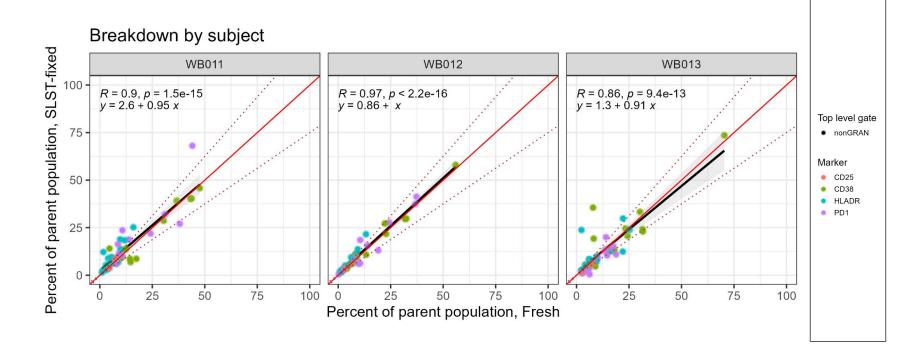
3. Reduce committed processing cost and expand analysis window Not sure which samples to analyze? No problem. Store samples at -80C until you're ready to decide which samples to analyze.

### FAQ

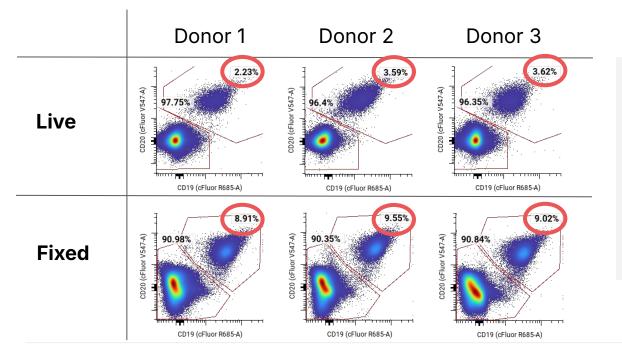
#### Subject level breakdown: phenotypic markers



#### Subject level breakdown: state markers



#### Where do we see differences? Total B-cells, for example.



Short answer: We do see consistent differences for certain immune population frequencies when comparing samples processed live versus fixed.

The biological relevance of this is an area of active investigation