You're in the right place.

To follow along at home, download our report at teiko.bio/spectral-flow-cytometry/#clia-validation-data

CLIA Validation for our 25-marker spectral flow test

High-parameter cytometry for clinical trials



Ramji Srinivasan Teiko CEO

What we tested

teiko.bio/spectral-flow-cytometry/#clia-validation-data

Spectral: Immune Profiling Markers

Our panel includes:

- All major immune cell population and subsets
- 5 functional state markers across subsets

T cells

Total T cells
CD4+ T cells
CD8+ T cells
Treg, activation
T cell subsets
Naive/Memory
Maturation
Maturation
gdT cells
NKT cells
Activation
Activation
Exhaustion

B cells

CD19	Total B cells
CD20	B cell subsets
CD27	Naive/Memory
IgD	B cell subsets
IgM	B cell subsets
CD25	Activation
HLA-DR	Antigen presentation

NK cells

CD56	NK cell subsets
CD16	NK cell subsets
CD38	Activation

Myeloid cells

CD14	Monocyte subsets
CD16	Monocyte subsets
CD11c	Monocytes, macrophages,
	DCs
CD123	pDC
CD141	cDC1
CD1C	cDC2
HLA-DR	Antigen presentation

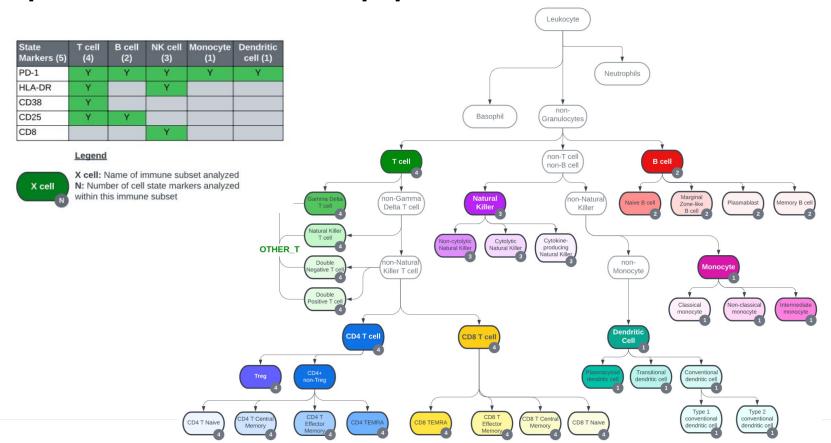
Granulocytes

CD123	Basophil
CD66b	Granulocytes

General

CD45	Total	immune	cells

Spectral Whole Blood: Cell populations and state markers



Tests used TokuKit fixed whole blood (WB) from three healthy human donors

Acquired samples from three healthy donors

TokuKit fixation at Teiko

Samples evaluated in test



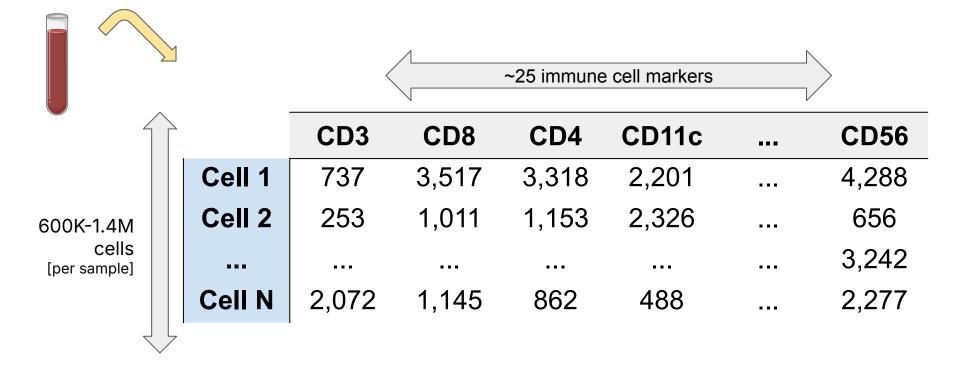












Each entry is a signal intensity

How do you reliably measure the immune state?



Is this test reproducible?

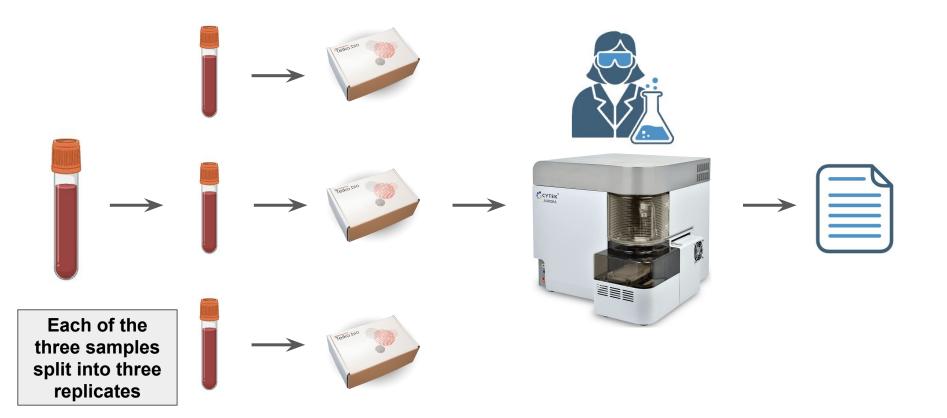
Measure	Meaning	Acceptance Criteria	Immune Populations Analyzed Per Donor*	Total Average Criteria Readout (%)	Total Pop. Below Criteria (%)
Intra-Run Precision	Same sample, same run	CV ≤ 20% for ≥95% of pop.	32	To come	
Inter-Run Precision	Same sample, different runs	CV ≤ 20% for ≥95% of pop.	32		

^{*}Only populations (pop.) with >100 median cells across three donors were included in analysis

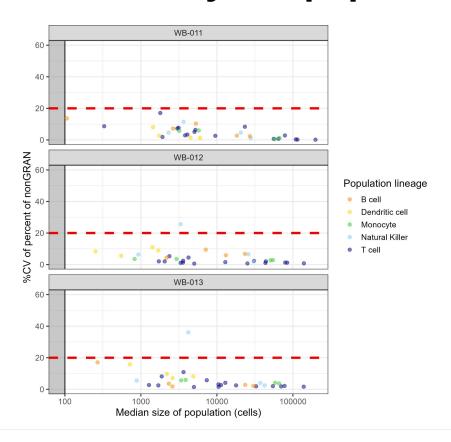
Intra-run

Intra-Run: Same sample, same run

Teiko.bio



Intra-Run by cell population size



Average %CV across all three donors:

4.76

Validation Results

Measure	Meaning	Acceptance Criteria	Immune Populations Analyzed Per Donor*	Total Average Criteria Readout (%)	Total Pop. Below Criteria (%)
Intra-Run Precision	Same sample, same run	CV ≤ 20% for ≥95% of pop.	32	4.76 %	100 % (32/32)
Inter-Run Precision	Same sample, different runs	CV ≤ 20% for ≥95% of pop.	32	To come	

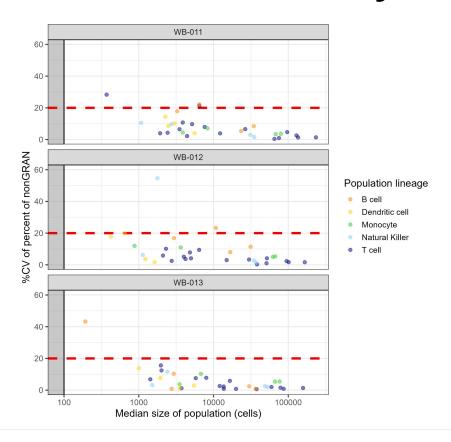
^{*}Only populations (pop.) with >100 median cells across three donors were included in analysis

Inter-run

Inter-Run: Same sample, different days, different operators

Day 1 Day 2 Day 3 Each of the three samples split into three replicates

Inter-Run Precision by cell population size



Average %CV across all three donors:

6.89

Measure	Meaning	Acceptance Criteria	Immune Populations Analyzed Per Donor*	Total Average Criteria Readout (%)	Total Pop. Below Criteria (%)
Intra-Run Precision	Same sample, same run	CV ≤ 20% for ≥95% of pop.	32	4.76 %	100 % (32/32)
Inter-Run Precision	Same sample, different runs	CV ≤ 20% for ≥95% of pop.	32	6.89 %	96.8 % (31/32)

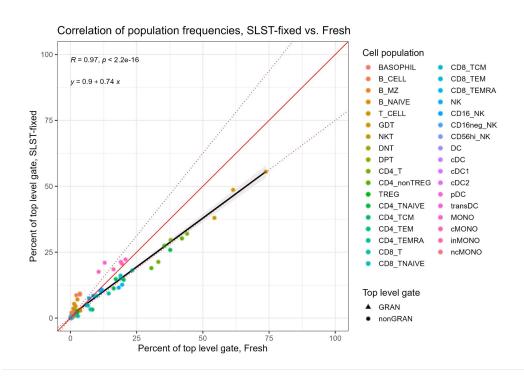
^{*}Only populations (pop.) with >100 median cells across three donors were included in analysis



Is this test comparable to fresh blood?



Check out our previous webinar:



Overall correlation of population frequencies between live and fixed sample processing: **0.97**

Why fix blood instead of running fresh?

1. 10-20X reduction of failure rate through centralization of flow analysis

No need for a flow lab in every country, city, clinical site. Reduce variability by

having all samples processed by the same facility.

2. Enables batching of samples

Conveniently store fixed samples until a whole set (for example all timepoints of an individual patient) is complete.

3. Reduce committed processing cost and expand analysis window
Not sure which samples to analyze? No problem. Store samples at -80C until
you're ready to decide which samples to analyze.

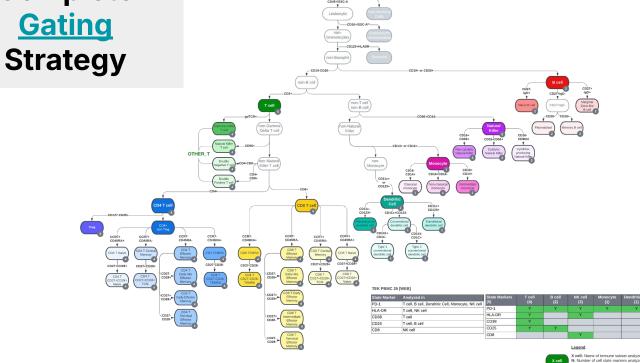
Read on

teiko.bio/spectral-flow-cytometry/#clia-validation-data

Appendix

Complete

Gating



Total Cells

X cell: Name of immune subset analyzed N: Number of cell state markers analyzed within this immune subset